

Comparative evaluation of phytochemical, in vitro antioxidant activities and elemental composition of the fruit, leaves, and stem bark of *Tetrapleura tetraptera*

Oyinkansade Yetunde Babalola¹, Ibraheem Oduola Lawal^{1*}, Idayat Adeola Akinwumi²

¹Biomedicinal Research Center, Forestry Research Institute of Nigeria, Ibadan, Nigeria; ² Department of Pharmacognosy, Faculty of Pharmacy, Lead City University, Ibadan, Nigeria

*Corresponding Authors: Ibrahim O. Lawal, Biomedicinal Research Center, Forestry Research Institute of Nigeria, Ibadan, 5054, Nigeria.

Submission Date: November 11th, 2023; Acceptance Date: December 19th, 2023; Publication Date: December 26th, 2023

Please cite this article as: Babalola O. Y., Lawal I. O., Akinwumi I. A. Comparative evaluation of phytochemical, in vitro antioxidant activities and elemental composition of the fruit, leaves, and stem bark of Tetrapleura tetraptera. *Functional Food Science* 2023; 3(12): 317-328. DOI: https://www.doi.org/10.31989/ffs.v3i12.1263

ABSTRACT

Introduction: Nowadays, medicinal plants are widely accepted as a better option than synthetic medications due to their lower cost and little to no side effects. The current study aimed to compare the elemental composition, phytochemical profile, and antioxidant efficacy of *n*-hexane extracts derived from *Tetrapleura tetraptera* fruit, stem bark, and leaves.

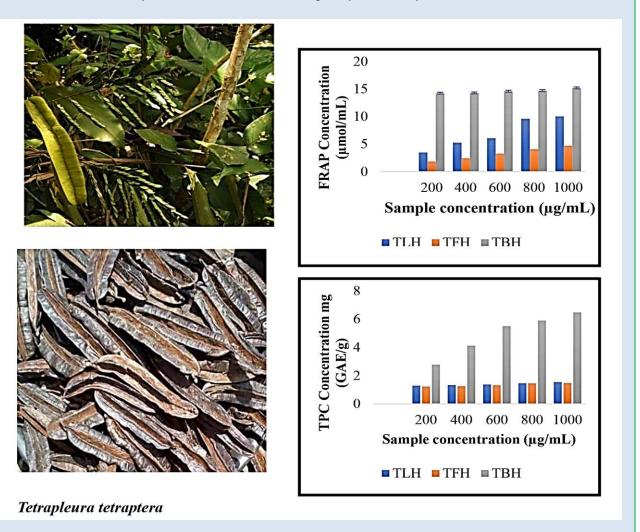
Methods: The fresh fruits, leaves, and stem bark of the plant were collected and dried. The dried samples were pulverized and exhaustively extracted with *n*-hexane. The extracts were screened for both qualitative and quantitative phytochemical constituents. Antioxidant parameters such as Ferric Reducing Antioxidant Property (FRAP) and 2,2-Diphenyl-1-picryl hydroxyl radical scavenging activity (DPPH) were also carried out. The elemental analysis of the various organs was also evaluated using standard protocols.

Results: The presence of alkaloids, flavonoids, tannins, saponins, terpenes, resins, and phenolswas revealed in the phytochemical data. The stem bark of *T. tetraptera* has higher concentrations of alkaloids and flavonoids than the leaf and fruit, according to a quantitative examination of the phytochemical elements of the plant. The stem bark had the highest Total Phenolic Content at $1000 \, \mu g/mL$, whereas the leaves had the lowest TPC at $200 \, \mu g/mL$.

At a concentration of 1000 μ g/mL, the stem bark exhibited the greatest activity of total flavonoid (TFC) in the plant extracts, followed by the fruits and leaves. At 1000 μ g/mL concentration, the FRAP test revealed the maximum activity in the stem bark. Generally speaking, when concentration rises, so do the TPC, TFC, and FRAP activities. Compared to the standard ascorbic acid (2.76 μ g/mL), stem bark exhibited the highest level of DPPH antioxidant activity, with an IC $_{50}$ of 13.34 μ g/mL. The elemental analysis revealed that the stem bark had the highest concentrations of Ca, Mg, and Fe, followed by the fruit and leaves, in that order.

Conclusion: This research result has established that the plant, *T tetraptera* can be standardized and offer an alternative eco-friendly drug for immunity building. The stem barks are underutilized, however, its phytochemical studies and other investigations pointed to its potential in the cosmeceutical industries and medicine.

Keywords: Antioxidant activity, ethnomedicine, ferric-reducing assay, medicinal plant



©FFC 2023. This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 License (http://creativecommons.org/licenses/by/4.0)

INTRODUCTION

People have employed medicinal plants and their extracts to treat a variety of ailments for thousands of years. Several different antitussives, antimalarial, antihypertensive, and analgesic medications are produced by these plants [1-2]. These therapeutic plants serve as significant starting points for the development of drugs that focus on a wide range of pharmacological targets, such as neurological diseases, cancer, malaria, and cardiovascular disease. In order to be categorized as a medicinal plant, a part of the plant or the entire plant must have medicinal qualities. Plant-based medications are cheaper and safer to use than conventional (synthetic) drugs, which often cause adverse side effects [3]. Plant derived medications include bioactive ingredients and extracts that have enormous potential for use in the development of unique and innovative methods for treating and preventing disease. These bioactive substances, which include steroids, cyanogenic glycosides, and essential oils, as well as tannins, flavonoids, alkaloids, phenols, phytates, and oxalates, can be employed as medications, be used as models for brand-new synthetic compounds, and taxonomic markers to identify novel compounds [2]. Plants play a crucial role in many scientific domains and are important to human survival. According to earlier studies, plant materials include several valuable natural compounds, such as coumarins, iridoids, alkaloids, and flavonoids [4]. The ethnopharmacology, phytochemistry, and biological activity of extensive collections of potential therapeutic plants from Africa and beyond have also been documented [5-7]. Compounds known as antioxidants prevent oxidation, a chemical process that can result in the production of free radicals. The regulation of physiological processes, cellular damage, and the etiology of brain disorders including dementia, stroke, and Parkinson's disease are all significantly influenced by free radicals [8, 9].

Tetrapleura tetraptera (Schumach. & Thonn.) Taub is a single-stemmed, sturdy, perennial tree that grows to a height of approximately 30 meters and belongs to the family Mimosaceae. The tree-like, strong, perennial plant has a single stem, thick, woody roots, and spreading branches in addition to dark green leaves. It has glabrous juvenile branches and a smooth or rough, grey-brown bark. The racemes of yellowish-pink flowers accompany the dark-brown, four-winged pods of the fruit. It is often found in tropical Africa's lowland forests. The fruit locally known as Aridan or prekese is made up of a luscious pulp and tiny, brownish-black seeds. It has an odor that is pleasant and distinctively pungent, which is linked to its ability to repel insects [10].

The goal of this study was to compare the antioxidant potential and the phytochemical and elemental composition of *n*-hexane extracts of leaves, stem bark, and fruit of *T. tetraptera*. Because medicinal plants are widely accepted today as being more effective than synthetic drugs and have fewer side effects, there is a great demand for natural antioxidants due to their lack of side effects. Many authors have studied the ethnomedicinal activity of the fruit extract of *T. tetraptera*, but little research has been done on the plant's stem bark.

METHODS

Chemicals and reagents: Analytical grade solvents such as methanol, n-hexane, chloroform, distilled water, ferric chloride, ammonia, Dragendorff's reagent, Mayer's reagent, sulphuric acid, glacial acetic acid, aluminum chloride, DPPH, potassium acetate (BDH Laboratory Supplies, Poole, England).

Plant collection and identification: Fresh fruits, leaves, and stem bark of *T. tetraptera* were collected from the herbal garden of the Forestry Research Institute of NigeriaThe plant samples were then carefully cleaned

using tap water and then rinsed using distilled water. The cleaned samples were air-dried on a rack for 10 days to reduce the water content and later transferred into a cabinet dryer at a temperature of 35-40°C for 10 min to further reduce the moisture. The dried plant samples were pulverized, using the laboratory milling machine. The powder obtained was filtered through a fine (2 mm) mesh sieve to get rid of any leftover material, it was kept in a sealed glass container for further analysis.

Extract preparation: Four hundred grams (400 g) of each powdered sample were exhaustively extracted with analytical grade n-hexane and were subjected to 24-hour continuous shaking using a laboratory shaker. The extracts were filtered using Whatman No. 42 (125 mm) filter paper. A rotary evaporator was used to concentrate the filtrate *in vacuo*. The extract was kept in a refrigerator at 4°C until utilized.

Qualitative analysis of phytochemical constituents:

Using established protocols, the extracts were evaluated for the presence of numerous secondary metabolites, including anthraquinones, alkaloids, tannins, saponins, flavonoids, cardiac glycosides, and steroids [11-12].

Quantitative analysis of phytochemical constituents:

Total phenolic content estimation (Folin–ciocalteu method): Standard procedures were used in order to determine the extracts' total phenolic content [13, 14]. 1 mL aliquots of 50–500 μg/mL ethanolic gallic acid solution were mixed with 5 mL of diluted folin–ciocalteu reagent and 4 mL of 7.5% sodium carbonate to prepare the standard calibration curve.

Total flavonoid content estimation: This was done according to the technique written about in an article by Elbouny et al. [14], in which the aluminum chloride colorimetric method was employed to determine the presence of flavonoids. To prepare the calibration curve,

a solution of quercetin was made in methanol at concentrations ranging from 20 to 100 µg/mL.

Ferric-reducing antioxidant property: The ferric-reducing property was evaluated by assessing the ability of extracts to reduce FeCl₃ solution. The test was performed according to standard procedures [15].

Determination of DPPH antioxidant activity: By using a slightly modified approach previously published the radical scavenging ability of the plant extracts against 2,2-Diphenyl-1-picryl hydroxyl radical (Sigma-Aldrich) was determined [14]. A positive control was employed, which was ascorbic acid. Based on the linear regression curve, the antioxidant activity of each sample was reported in terms of IC₅₀, or the micromolar concentration needed to prevent the generation of DPPH radicals by 50%. The following formula was used to compute the radical scavenging activity.

% inhibition = {[Ab-Aa] /Ab} x 100, where Aa represents the extract's absorption and Ab represents the blank sample's absorption.

Elemental Analysis: The elemental composition of the fruits, leaves, and stem bark of *T. tetraptera* was carried out using a standard analytical method [16], where the powdered samples were digested using nitric and perchloric acids. Potassium and Sodium were assessed using a Flame photometer, while magnesium, calcium, iron, copper, manganese, zinc, and phosphorus were determined from the filtered aliquots using an Atomic Absorption Spectrophotometer (Buck Scientific; 210VGP Model) (SearchTech British; FP640 Model). Each analysis was performed in triplicates.

Statistical analysis: Microsoft Excel software (2010) was used for statistical analysis. Three duplicates of each analysis were carried out.

RESULTS

Qualitative phytochemical screening: The results of the phytochemical screening revealed the presence of phenols, steroids, alkaloids, flavonoids, anthraquinones, tannins, and terpenoids except for saponin and tannin which were present only in the stem bark but absent in

the fruit and leaves of the plant and cardiac glycoside which was completely absent in all (Table 1). The presence of these secondary metabolites qualifies the plant as a medicinal plant.

Table 1. Qualitative analysis of phytochemical constituents of *T. tetraptera*

Phytochemical constituents	Test	Fruit	Leaf	Stem bark
Alkaloid	Dragendorff	+	+	+
	Mayer	+	+	+
	Wagner	++	++	++
Saponins	Frothing	-	-	+
Glycoside	Keller-Killian	-	-	-
Flavonoids	Ammonia/H₂SO ₄	+	+	+
	Aluminum solution	+	+	+
	Ethyl acetate/Ammonia	++	++	++
Anthraquinones	Borntrager	+	+	+
Terpenoids	Salkwoski's	++	+	++
Tannins	Ferric chloride	-	-	++
Steroids	Liebermann-Burchard	++	++	+

Interpretation: + Present; ++ Abundant; -ve Absent.

Quantitative phytochemical screening: The quantitative analysis of phytochemical constituents of *T. tetraptera* revealed the highest concentration of alkaloids and

flavonoids in the stem bark as compared to the leaf and fruit (Table 2).

 Table 2. Quantitative analysis of phytochemical constituents of T. tetraptera

Phytochemical	Leaf (%)	Fruit (%)	Stem bark (%)
constituents			
Alkaloids	12.7	20	33.5
Flavonoids	19	26.7	52.3
Saponin			9.1

Total phenolic content (TPC): At 1000 $\mu g/mL$ concentration, the stem bark (TBH) had the greatest Total Phenolic Content (6.47 mg GAE/g), whereas at 200

 $\mu g/mL$, the leaves (TLH) had the lowest TPC (1.28 mg GAE/g) (Fig. 1).

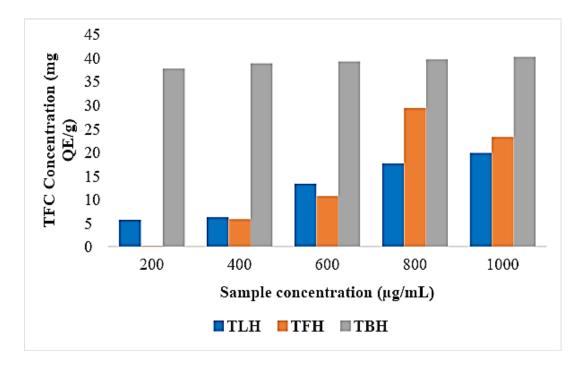


Figure 1. Total phenolic content (TPC) of samples

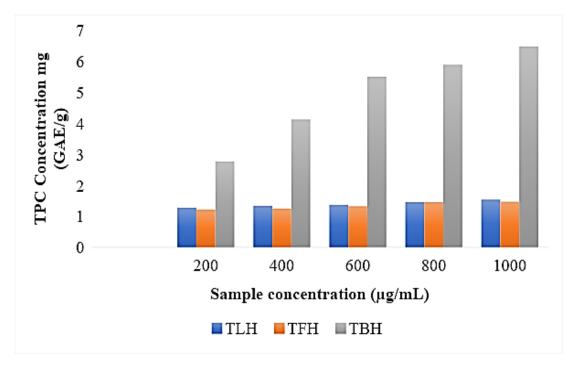


Figure 2. Total flavonoid content (TFC) of sample

Total flavonoid content (TFC): The total flavonoid (TFC) of the plant extracts showed the highest activity in the

stem bark (TBH) (40.15 mg QE/g), followed by the fruits (TFH) (23.17 mg QE/g) and the leaves (TLH) (19.82 mg QE/g) at1000 μ g/mL concentration (Figure 2).

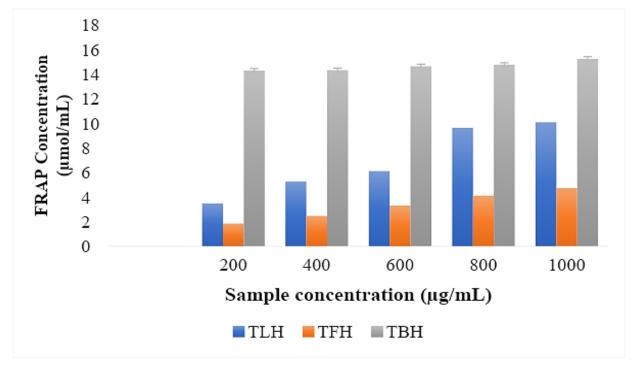


Figure 3. Ferric reducing antioxidant power (FRAP) of samples.

Ferric reducing antioxidant power assay (FRAP): The FRAP result showed the highest activity in the stem bark (TBH) (15.22 μ mol/mL) at 1000 μ g/mL concentration (Fig. 3). In general, the TPC, TFC, and FRAP activities increase as the concentration increases (Fig. 1-3).

Table 3 shows the DPPH free radical scavenging activity of *T. tetraptera* extracts. The stem bark was found to be most active with IC $_{50}$ of 13.34 $\mu g/mL$ when compared with the standard, ascorbic acid 2.76 $\mu g/mL$.

Table 3. DPPH radical scavenging % inhibition of *T. tetraptera*

Plant parts	% Inhibition	
Ascorbic acid	2.76 ± 0.07	
Stem bark	13.34 ± 1.43	
Leaves	449.59 ± 149.70	
Fruit	7815 ± 309.01	

Values represent Mean ± SEM.

Elemental analysis: The result of the elemental analysis carried out using atomic absorption spectrophotometry

is presented in Table 4. The fact that Ca, Mg, K, Cu, Fe, and Zn are present indicates their role as necessary

nutritional components. The highest Ca, Mg, and Fe concentrations were found in the stem bark followed by the fruit and the leaves respectively. This shows that

different parts of plants take up and absorb various nutrients from the soil in varying amounts.

Table 4. Mineral composition of *T. tetraptera* stems, bark, leaves and fruits on when dry.

Elements	Quantity (mg/100g)		
	Stem bark	Leaves	Fruit
Calcium	3.55±0.56	0.43±22.29	0.45±12.61
Magnesium	1.81±0.83	0.99±0.04	0.71±0.48
Copper	0.02±11.11	0.19±1.37	0.09±7.12
Iron	1.08±11.70	4.02±1.94	2.37±1.84
Zinc	0.28±4.88	0.32 ±1.79	0.27 ±7.23
Potassium	4.89±00	4.89±00	4.89+00

Values represent Mean ± SEM

DISCUSSION

Alkaloids are basic and naturally occurring organic compounds with at least one nitrogen atom. This is a vastly diversified group of naturally occurring compounds generated through secondary metabolism (does not include nitrogen in an amide bond or peptide). They are frequently isolated from plants, although various mammals, insects, marine invertebrates, and other microbes have also been shown to contain them [17, 18]. Our present study established a comparison evaluation of the phytochemical, in vitro antioxidant activities and elemental composition of the various parts of T. tetraptera. The fruit, stem bark, and leaf extracts of the plant, T. tetraptera, all contain alkaloids, flavonoids, anthraquinones, terpenoids and steroids which indicates that the plant has therapeutic qualities. Saponins play a critical role in the stimulation of mucous membrane defensive factors thereby exerting its defensive factors in gastric ulceration [19-20].

Early studies have shown that saponins affect fish, Oncomelania snails, and insects biologically, in addition to having hemolytic and antibacterial properties. It has been established that saponins are risk-free, environmentally beneficial, and microbially degradable [21-22].

The culinary and medicinal industries make extensive use of tannins' antioxidant qualities. Tannins have received a lot of attention due to their antioxidant properties, which include fighting cancer, osteoporosis, and cardiovascular disease [23]. Several biological activities of tannins have been reported [24]. These include antibacterial [25-26], antiviral [27-28], antiparasitic [29-30], anti-inflammatory [31-32], antidiarrheal [33-34], and antioxidant [23].

The present antioxidant activity result showed that *T. tetraptera* is rich in antioxidants which may protect the human cells against free radicals and thereby play a significant role in the treatment of heart diseases, cancer,

and other diseases. The stem bark had the highest concentrations of TPC, TFC, and FRAP.

This study reported that the *T. tetraptera* plant, especially the stem bark possesses the highest concentrations of tannins flavonoids, phenolics, and FRAP. It has been shown that flavonoids and phenols have antiplatelet, anticancer, anti-inflammatory, and antioxidant properties [35].

Many fruits and vegetables, including tomatoes, berries, and grapes, contain phenolic compounds. By lowering the chance of developing metabolic diseases like type 2 diabetes mellitus, phenolic compounds can improve one's health [36]. Numerous research has examined their impact on antioxidant, anti-inflammatory, anti-aging, and antiproliferative activities [37-39].

Flavonoids have been associated with several health-promoting advantages, making them crucial for a broad variety of applications in nutraceuticals, pharmacology, medicine, and cosmetics. This is because they have potent anti-oxidative, anti-inflammatory, antimutagenic, antibacterial, anti-carcinogenic, and vascular properties. They also have the therapeutic capacity to alter important cellular enzyme activities and scavenge free radicals [40].

The origin, development, and advancement of tumors have been demonstrated to be inhibited by flavonoids, which are also known to have antioxidant properties [41]. There have been reports linking flavonoid consumption to a decrease in coronary heart disease [42]. Other biological activities that flavonoids perform in addition to acting as antioxidants include defense against platelet aggregation, free radicals, bacteria, viruses, hepatotoxins, inflammation, cancers, ulcers, and allergies [43]. Both humans and plants use phenols as antioxidants [44]. Haslam [45] draws attention to the growing interest in the possibility of treating illnesses by simply increasing dietary

consumption of foods abundant in plant phenolics including tannins and flavonoids, as well as antioxidants like vitamins E, C, -carotene, and carotenoids. The ability of the *n*-hexane extract of *T. tetraptera* bark to scavenge DPPH radicals indicates its antioxidant activity.

In addition, the elemental analysis showed an abundance of mineral elements than the leaves and fruits. The mineral elements Calcium, Magnesium, and Potassium present in abundant in the stem bark showed that *T. tetrapleura* could play a significant role in the body's metabolism. This edible species can be made in supplements because trace elements supplements are beneficial in the treatment of type 2 diabetes mellitus, prevention of hypoglycemia, corticosteroid-induced hyperglycemia, molybdenum deficiency, and others.

In addition to providing food for both plants and animals, mineral elements also play other significant roles in the ecosystem. It has long been established that inorganic chemical elements are crucial for nourishment and serve as significant structural elements in cellular functions [46]. When used medicinally for a therapeutic effect, the amount of trace elements and active ingredients in medicinal plants have a significant influence on the body's metabolism [47].

The elements Fe⁺², K⁺, Mg⁺², Na⁺, Ca⁺², Co⁺², Mn⁺², Zn⁺², and Cu⁺² is considered essential, and Ni⁺² and Cr⁺³ is possibly essential, and Cd⁺², Pd⁺⁴, and Li⁺¹ is considered non-essential elements of the human body.

Zinc is an essential micronutrient, which makes it an important component of the plant life cycle [48]. Copper is another important micronutrient for plants [49]. It is involved in several metabolic activities, including the scavenging of superoxide, hormone signaling, electron transport during photosynthetic reactions, mitochondrial respiration, and cell wall metabolism. One of the critical micronutrients for plant growth and development is iron. The synthesis of DNA, chlorophyll, respiration, photosynthesis, hormones, and nitrogen-fixing are just a

few of the biological processes in which it takes part [50-51].

CONCLUSION

In addition to being used for timber, *T. tetraptera* may also be used in traditional medicine. Most of its leaf components, including the seeds, fruit, and leaves, have been used in traditional medicine in well-known ways, while the stem bark has received less attention from pharmaceutical and phytochemical companies.

This research has established through the investigation of this plant's phytochemistry and antioxidant properties that *T. tetraptera* has industrial potential, especially for use in pharmaceuticals. The efficacy of the fruit and stem bark extracts of *T. tetraptera* in its bioactivities may be attributed to its potential antioxidant activity. Based on the findings, stem bark was more effective as an antioxidant. The stem bark of this plant should not be neglected as could be explored for further biological studies and chemical evaluation. It is recommended that further research be done on the biological actions of active ingredients and isolation.

List of abbreviations: Ca: Calcium, Mg: Magnesium, K: Potassium, Cu: Copper, Fe: Iron, Zn: Zinc, TPC: Total Phenolic Content, TFC: Total flavonoid content, DPPH: 2,2-diphenyl-1-picrylhydrazyl, FRAP: Ferric reducing antioxidant power.

Competing interests: The authors declare that there are no competing interests.

Authors' contributions: All authors performed the experiment and data analyses. OYB wrote the first draft of the manuscript. IOL and IAA reviewed and edited the manuscript. OYB and IOL contributed to the conceptualization, project administration, and financial support. IOL guided the experiment and revised the

manuscript. All authors approved the final version of the manuscript.

Acknowledgements and Funding: This research is selffunded by the authors.

REFERENCES

- Mohan N, Jassal PS, Singh RP: Comparative in vitro and in vivo study of antioxidants and phytochemical content in Bacopa monnieri. Recent Res Sci Technol 2018, 3(9):78-83.
 DOI: https://doi.org/10.1186/s12906-019-2534-4
- Nsofor WN, Nwaoguikpe RN, Ujowundu FN, Keke CO, Uba MT, Edom CV: Phytochemical, GC-MS, FTIR and Amino acid profile of methanol extract of *Tetrapleura tetraptera* fruit. J Drug Del Ther 2023, 13(2):61-69.

DOI: https://doi.org/10.22270/jddt.v13i2.5739

- Ngozi Al: Phytomedicines and Nutraceuticals: Alternative Therapeutics for Sickle Cell Anemia", The Sci World J 2013, 12pp. DOI: https://doi.org/10.1155/2013/269659
- Mohammadhosseini M, Venditti A, Sarker SD, Nahar L, Akbarzadeh A: The genus Ferula: Ethnobotany, phytochemistry and bioactivities - a review. Ind Crops Prod 2019, 129:350-394.

DOI: https://doi.org/10.1016/j.indcrop.2018.12.012

- Nalawade AS, Gurav RV, Patil AR, Patwekar M, Patwekar F:
 A comprehensive review on morphological, genetic and phytochemical diversity, breeding and bioprospecting studies of genus *Chlorophytum* Ker Gawl. from India. Trends Phytochem Res 2022, 6(1):19-45.
 - DOI: https://doi.org/10.30495/tpr.2022.1949493.1238
- Olaoluwa O, Taiwo O, Nahar L, Sarker SD: Ethnopharmacology, phytochemistry and biological activities of selected African species of the genus *Ficus*. Trends Phytochem Res 2022, 6(1):46-69.
 - DOI: https://doi.org/10.30495/tpr.2022.1939285.1219
- Akinwumi IA, Sonibare MA: Sphenocentrum jollyanum Pierre (Menispermaceae): From traditional medicine to pharmacological activity and chemical constituents. Trends Phytochem Res 2022, 6(4):301-313.

DOI: https://doi.org/10.30495/tpr.2022.1961991.1268

 Lin TK, Der CS, Lin KJ, Chuang YC: Seizure-induced oxidative stress in status epilepticus: is antioxidant beneficial? Antioxidants 2020, 9:1–24.

DOI: https://doi.org/10.3390/antiox9111029

 Oladejo CO, Ogundele OO, Adeoti AR, Atilola JR, Olaleye MT, Akinmoladun AC: Tetrapleura tetraptera curtails oxidative and pro-inflammatory biochemical events in the lithiumpilocarpine model of status epilepticus. Adv Trad Med 2022, 1-12.

DOI: https://doi.org/10.1007/s13596-022-00675-x

 Ojewole JA, Adewunmi CO: Anti-inflammatory and hypoglycaemic effects of *Tetrapleura tetraptera* (Taub) (Fabaceae) fruits aqueous extract in rats. J Ethnopharmacol 2004, 95:177-182.

DOI: https://doi.org/10.1016/j.jep.2004.06.026

- Sofowora A: Medicinal Plants and Traditional Medicine in Africa. 2nd Ed. Sunshine House, Ibadan, Nigeria: Spectrum Books Ltd; Screening Plants for Bioactive Agents. 1993, pp. 134-156.
- Trease GE, Evans WC: Pharmacognosy. 15th Ed. London: Saunders Publishers, 2002, pp. 42-44, 221-229, 246-249, 304-306, 331-332, 391-393.
- Khatoon M, Islam E, Islam R, Rahman AA, Khurshid AHM, Khondkan P: Estimation of Total Phenol and *In vitro* antioxidant activity of *Albizia procera* leaves. Springer 2013, 3:21-33. DOI: https://doi.org/10.1186/1756-0500-6-121
- 14. Elbouny H, Ouahzizi B, Khouya T, Bakali AH, Sellam K, Alem C: Total phenolic compounds, and antioxidant activity of eight Moroccan *Thymus species*: a comparison with *Thymus vulgaris*. Herba Pol 2023, 69(1).

DOI: https://doi.org/10.5604/01.3001.0016.2981

15. Benzie I, Devaki M: The ferric reducing/antioxidant power (FRAP) assay for non-enzymatic antioxidant capacity: concepts, procedures, limitations, and applications: recent trends and applications. In: Apak R, Capanoglu E, Shahidi F (eds). Measurement of Antioxidant Activity and Capacity: Recent Trends and Applications. Wiley 2017.

DOI: https://doi.org/10.1002/9781119135388.ch5

- Association of Official Analytical Chemists. Official Methods of Analysis of the Association of Official Analytical Chemists International, 2010, 18th edition. Washington DC, USA.
- **17.** Bribi N: Pharmacological activity of alkaloids: a review. Asian J Bot 2018, 1. DOI: https://doi.org/10.63019/ajb.v1i2.467
- Gutierrez-Grijalva EP, Lopez-Martinez LX, Contreras-Angulo LA, Elizalde-Romero CA, Heredia JB: Plant Alkaloids: Structures and Bioactive Properties. Springer Nature Singapore Pte Ltd. M. K. Swamy (ed.), Plant-derived Bioactives 2020.

DOI: https://doi.org/10.1007/978-981-15-2361-8 5

 Choudhary MK, Bodakhe SH, Gupta SK: Assessment of the antiulcer potential of *Moringa oleifera* root-bark extract in rats. J Acupunct Meridian Stud 2013, 6:214–220.

DOI: https://doi.org/10.1016/j.jams.2013.07.003

- Akinwumi IA, Sonibare MA, Salami AT: Gastroprotective Effect of Methanol Extracts of Sphenocentrum jollyanum Pierre and Curculigo pilosa (Schumach. & Thonn.) Engl. in Wistar Rats. Trop J Nat Prod Res 2022, 6(4):637-644.
- Akagi M, Fukuishi N, Kan T, Sagesaka YM, Akagi R: Antiallergic effect of tealeaf saponin (TLS) from tea leaves (*Camellia sinensis* var. *sinensis*). Biol Pharm Bull 1997, 20:565-567. DOI: https://doi.org/10.1248/bpb.20.565
- Cui C, Zong J, Sun Y, Zhang L, Ho CT, Wan X, Hou R: Triterpenoid saponins from the genus *Camellia*: structures, biological activities, and molecular simulation for structure—activity relationship. Food Funct 2018, 9(6):3069–3091. DOI: https://doi.org/10.1039/c8fo00755a23
- Squillaro T, Cimini A, Peluso G, Giordano A, Melone MAB: Nano-delivery systems for encapsulation of dietary polyphenols: an experimental approach for neurodegenerative diseases and brain tumors. Biochem Pharmacol 2018, 154:303–317.

DOI: https://doi.org/10.1016/j.bcp.2018.05.016

24. Tong Z, He W, Fan X, Guo A: Biological Function of Plant Tannin and Its Application in Animal Health. Front Vet Sci 2022, 8:803657.

DOI: https://doi.org/10.3389/fvets.2021.803657

Jamroz D, Wiliczkiewicz A, Skorupinska J, Orda J, Kuryszko J,
Tschirch H: Effect of sweet chestnut tannin (SCT) on the
performance, microbial status of intestine and histological
characteristics of the intestine wall in chickens. Br Poult Sci
50:687–99.

DOI: https://doi.org/10.1080/00071660903191059

 Biagia G, Cipollini I, Paulicks BR, Roth FX: Effect of tannins on growth performance and intestinal ecosystem in weaned piglets. Arch Anim Nutr 2010, 64:121–135.

DOI: https://doi.org/10.1080/17450390903461584

- Liu S, Chen R, Hagedorn CH: Tannic acid inhibits hepatitis C virus entry into Huh 7.5 cells. PLoS ONE 2015, 10: e0131358.
 DOI: https://doi.org/10.1371/journal.pone.0131358
- Liu C, Cai D, Zhang L, Tang W, Yan R, Guo H: Identification of hydrolyzable tannins (punicalagin, punicalin and geraniin) as novel inhibitors of hepatitis B virus covalently closed circular DNA. Antivir Res 2016, 134:97–107.

DOI: https://doi.org/10.1016/j.antiviral.2016.08.026

 Oliveira LM, Bevilaqua CM, Macedo IT, Morais SM, Monteiro MV, Campello CC: Effect of six tropical tanniferous plant extracts on larval escheatment of *Haemonchus contortus*. Rev Bras Parasitol Vet 2011, 20:155–160.

DOI: https://doi.org/10.1590/s1984-29612011000200011

 Molan AL: Effect of purified condensed tannins from pine bark on larval motility, egg hatching and larval development of *Teladorsagia circumcincta* and *Trichostrongylus* colubriformis (Nematoda: Trichostrongylidae). Folia Parasitol (Praha) 2014, 61:371–376.

DOI: https://doi.org/10.14411/fp.2014.036

 Pallares V, Cedo L, Castell-Auvi A, Pinent M, Ardevol A, Arola L: Effects of grape seed procyanidin extract over low-grade chronic inflammation of obese Zucker fa/fa rats. Food Res Int 2013, 53:319–324.

DOI: https://doi.org/10.1016/j.foodres.2013.05.006

32. Park M, Cho H, Jung H, Lee H, Hwang KT: Antioxidant and anti-inflammatory activities of tannin fraction of the extract from black raspberry seeds compared to grape seeds. J Food Biochem 2014, 38:259–270.

DOI: https://doi.org/10.1111/jfbc.12044

- Bonelli F, Turini L, Sarri G, Serra A, Buccioni A, Mele M: Oral administration of chestnut tannins to reduce the duration of neonatal calf diarrhea. BMC Vet Res 2018, 14:227.
 - DOI: https://doi.org/10.1186/s12917-018-1549-2
- 34. Girard M, Thanner S, Pradervand N, Hu D, Ollagnier C, Bee G: Hydrolysable chestnut tannins for reduction of postweaning diarrhea: efficacy on an experimental ETEC F4 model. PLoS ONE 2018, 13: e0197878.

DOI: https://doi.org/10.1371/journal.pone.0197878

- 35. Pal D, Verma P: Flavonoids: a powerful and abundant source of antioxidants. Int J Pharm Pharm Sci 2013, 5:97–110.
- Gasmi A, Mujawdiya PK, Noor S, Lysiuk R, Darmohray R,
 Piscopo S: Polyphenols in metabolic diseases. Molecules 2022, 27(19):6280.

DOI: https://doi.org/10.3390/molecules27196280

- Lee JH, Park J, Shin DW: The molecular mechanism of polyphenols with anti-aging activity in aged human dermal fibroblasts. Molecules 2022, 27(14): 4351. DOI: https://doi.org/10.3390/molecules27144351
- Bie J, Sepodes B., Fernandes PC, Ribeiro MH: Polyphenols in health and disease: Gut microbiota, bioaccessibility, and bioavailability. Compounds 2023, 3(1): 40–72.
 DOI: https://doi.org/10.3390/compounds3010005
- Elgadir MA, Chigurupati S, Mariod AA: Selected potential pharmaceutical and medical benefits of phenolic compounds: Recent advances. Functional Food Science

DOI: https://www.doi.org/10.31989/ffs.v3i7.1118

2023, 3(7):108-128.

 Karak P: Biological activities of flavonoids: An overview. Int J Pharm Sci Res 2019, 210(4):1567-1574. DOI: https://doi.org/10.13040/JJPSR.0975-8232.10(4).1567-74

- 41. Kim SY, Kim JH, Kim SK, Oh MJ, Jung MY: Antioxidant activities of selected oriental herb extracts. J Am Oil Chem Soc 1994, 71(6):633–640.
- Hertog MGL, Feskens EJM, Hollman PCH, Katan JB, Kromhout D: Dietary antioxidant flavonoids and risk of coronary heart disease: the Zutphen Elderly Study. The Lancet 1993, 342(8878):1007–1011.
 DOI: https://doi.org/10.1016/0140-6736(93)92876-u
- 43. Barakat MZ, Shahab SK, Darwin N, Zahemy EI:

 Determination of ascorbic acid from plants. Anal Biochem
 1993, 53:225–245.
- 44. Dillard CJ, German JB: Phytochemicals: nutraceuticals and human health. J Sci Food Agric 2000, 80(12):1744–1756. DOI:<a href="http://dx.doi.org/10.1002/1097-0010(20000915)80:12<1744::AID-JSFA725>3.0.CO;2-W">http://dx.doi.org/10.1002/1097-0010(20000915)80:12<1744::AID-JSFA725>3.0.CO;2-W
- Haslam E: Practical Polyphenolics: From Structure to Molecular Recognition and Physiological Action, Cambridge University Press 1998, Cambridge, UK.
- 46. Willis JA, Scott RS, Brown LJ: Islet cell antibodies and antibodies against glutamic acid decarboxylase in newly diagnosed adult-onset diabetes mellitus. Diabetes Res Clin Pract 1996, 33:89–97.

DOI: https://doi.org/10.1016/0168-8227(96)01281-8

- Rasheed MN: Trace elements in some wild plants from the shores of the high dam lake and the adjacent desert as determined by atomic absorption spectroscopy. J Arid Environ 1995, 29:185-197.
 - DOI: https://doi.org/10.1016/S0140-1963(05)80089-8
- 48. Horeth S, Detterbeck A, Ahmadi H, Pongrac P, Clemens S: Molecular analysis of-within plant Zn mobility in a metal hyperaccumulator and a crop model system. In OF Abstracts 2015 (p. 22).
- Adrees MAS, Rizwan M, Ibrahim M, Abbas F, Farid M: The effect of excess copper on growth and physiology of important food crops: a review. Environ Sci Pollut Res 2015, 22(11):8148–8162.

DOI: https://doi.org/10.1007/s11356-015-4496-5

- Tavsan Z, Kayali HA: The effect of iron and copper as an essential nutrient on mitochondrial electron transport system and lipid peroxidation in *Trichoderma harzianum*.
 Appl Biochem Biotechnol 2013, 170(7):1665–1675. DOI: https://doi.org/10.1007/s12010-013-0273-4
- Vatansever R, Filiz E, Ozyigit II: Genome-wide analysis of iron-regulated transporter 1 (IRT1) genes in plants. Hortic Environ Biotechnol 2015, 56(4):516–523.

DOI: https://doi.org/10.1007/s13580-015-0014-4